

New Insights Into the Pathogenesis of Cystic Fibrosis

Pivotal Role of Glutathione System Dysfunction and Implications for Therapy

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Abstract

The cystic fibrosis transmembrane regulator (CFTR) should no longer be viewed primarily as a 'chloride channel' but recognized as a channel that also controls the efflux of other physiologically important anions, such as glutathione (GSH) and bicarbonate. More effective approaches to cystic fibrosis treatment may result from this reconceptualization of the CFTR by researchers and clinicians. For example, oxidant damage in cystic fibrosis has been assumed to be a significant part of the pathophysiology of the disease. Generally speaking, antioxidant status in cystic fibrosis is compromised. However, until recently this was seen as secondary to the excessive chemoattraction of neutrophils in this disease caused by mutation of the CFTR protein, leading to a high oxidant burden. New findings suggest that the cystic fibrosis mutations in fact cause a primary dysfunction in the system of one of the body's most important antioxidant and immune-signaling substances: the reduced GSH system. Cystic fibrosis mutations significantly decrease GSH efflux from cells without redundant channels to the CFTR; this leads to deficiency of GSH in the epithelial lining fluid of the lung, as well as in other compartments, including immune system cells and the gastrointestinal tract. This deficiency is exaggerated over time as the higher-than-normal oxidant burden of cystic fibrosis leads to successively larger decrements in GSH without the normal opportunity to fully recover physiologic levels. This GSH system dysfunction may be the trigger for initial depletion of other antioxidants and may also play a role in initiating the over-inflammation characteristic of cystic fibrosis. Proper GSH system functioning also affects immune system competence and mucus viscosity, both of relevance to cystic fibrosis pathophysiology. In a way, cystic fibrosis may be thought of as the first identified disease with GSH system dysfunction.

This overview provides a review of the most pertinent recent research findings in this area. Exogenous augmentation of GSH in the lung epithelial lining fluid is possible, and therapeutic approaches include administration of aerosolized buffered GSH, intravenous GSH, and oral GSH. However, it is important to remember that the pathophysiology of cystic fibrosis is multifactorial, and rectification of GSH system dysfunction in patients with cystic fibrosis will not eliminate all harmful effects of the disease. The promising results of two clinical trials of aerosolized buffered GSH in cystic fibrosis patients have been published or accepted for publication at the time of this writing. GSH depletion in lung epithelial lining fluid has also been noted in other respiratory diseases such as COPD, idiopathic pulmonary fibrosis, and adult respiratory distress syndrome, and therapies to augment GSH may also be contemplated in these diseases.

Cystic fibrosis disease is the result of a mutation of the cystic fibrosis transmembrane regulator (CFTR) protein, resulting in missing or defective cellular anion efflux channels in epithelial cells. Most cystic fibrosis patients ultimately die of respiratory failure, as a result of deterioration in pulmonary function. There are several causes for deterioration in lung function, including

colonization of the lungs by bacteria and fungi and auto-destruction of lung tissue by excessive inflammation even in the absence of pathogen challenge (and accelerated by pathogen challenge when it occurs). The excessive inflammation has been linked to the cystic fibrosis mutation itself and is associated with greater than normal chemoattraction for neutrophils as a result of higher

constitutive levels of, for example, interleukin (IL)-8.^[1-8] The oxidant burden caused by such excessive inflammation appears to overwhelm natural antioxidant defenses. The resulting damage to lung tissue allows for greater adhesion of pathogens,^[9] in addition to reducing lung function directly.^[10]

New research findings, however, suggest that the traditional view of disease pathogenesis needs some modification. It now appears that cystic fibrosis mutations cause a primary dysfunction in one of the most important antioxidant and immune-signaling substances: the reduced glutathione (GSH) system. Dysfunction in the GSH system may be the catalyst for initial depletion of other antioxidants and may play a role in priming and perpetuating excessive inflammation characteristic of cystic fibrosis. Nevertheless, it is important to remember that cystic fibrosis pathophysiology is multifactorial and that not all cystic fibrosis disease manifestations can be linked to GSH system dysfunction.

This modification of the traditional view is in line with cutting-edge cystic fibrosis research, which suggests that the sole focus on the CFTR channel as a chloride efflux channel has obscured other significant functions associated with it. Current research now views the CFTR channel as, at the very least, a chloride, bicarbonate, and GSH efflux channel. Undoubtedly this list will be expanded in the future. This new and fundamental shift in the way the CFTR channel is viewed may allow for the development of innovative and effective therapies for cystic fibrosis. In this article, we will focus on the CFTR as a GSH efflux channel and discuss its significance for the respiratory system. It is worth noting that preliminary studies of the role of bicarbonate secretion in the lung have shown that the lack of functional CFTR may result in a decrease in the pH of the cystic fibrosis lung, which may also have pathologic consequences;^[11,12] however, this finding is disputed elsewhere.^[13-15] The newly understood role of CFTR as a GSH and bicarbonate efflux channel also has important implications for gastrointestinal complications associated with cystic fibrosis. In this article, discussion will be confined to the effects of CFTR mutation on the respiratory system only.

1. Understanding the Role of the Glutathione (GSH) System in Normal Lung Health

Virtually all cells of the body produce thiol-reduced GSH from the three amino acids glutamine, glycine, and cysteine. Cysteine serves as the rate-limiting amino acid for GSH production. It is estimated that an adult male produces approximately 10g of GSH per day. Not only is GSH present in the cells of the body, it also bathes the extracellular spaces of the body, with high extracellular levels in organ systems that come in contact with the oxidant-rich atmosphere, such as the cornea and the lung. One of the most

important roles of GSH, then, is to act as a water-soluble antioxidant. Not only can it neutralize oxidants through an enzymatic pathway utilizing GSH peroxidase, it is also capable of neutralizing oxidants directly without the use of an enzymatic pathway.^[16] In cystic fibrosis, however, epithelial cells still produce GSH normally, but one result is significant impairment of the ability of cells that do not possess a channel redundant to the CFTR to efflux GSH to fulfill its functions in the extracellular milieu.

Each antioxidant system – fat-soluble, water-soluble, and enzymatic – protects the cell within its own sphere of action. Some systems operate within the cytosol, which others operate at the cell membrane or are active in the extracellular milieu. GSH operates both within the cytosol and in the extracellular milieu. In these compartments, it is capable of directly reducing oxidants and it also reacts with GSH peroxidase, located in the cell membrane, to neutralize oxidants. GSH is replenished in two ways: by interaction of GSH disulfide (GSSG) with the enzyme GSH reductase and by synthesis of GSH within the cell (i.e. *de novo* or after cleavage of extracellular GSH and transport of component amino acids back into the cell). Circulation of GSH effluxed from cells throughout the body may allow for higher levels of GSH particularly in the extracellular compartments, such as the lung.

The various antioxidant systems are interdependent, to one degree or another, for proper function. The crippling of one system leads to decreased protection by other antioxidant systems. Without GSH, as we have seen, GSH peroxidase is unable to function as an antioxidant.^[17] Antioxidant systems such as GSH, ascorbic acid, tocopherol, and ubiquinol-10^[18,19] are interdependent, and normal levels of each in reduced form are required to maintain normal levels of the others in reduced form.^[20] In addition, GSH deficiency is linked to decreased activity of catalase and superoxide dismutase.^[20] GSH deficiency also taxes the fat-soluble antioxidant systems by permitting greater levels of lipid peroxidation, yielding damaging metabolites.^[20-27] This genetic chink in the antioxidant armor of cystic fibrosis patients predisposes them to have successively larger decrements in antioxidant protection over time, as other antioxidants are consumed in greater quantities or left unused as a result of impaired GSH efflux.

Furthermore, recent studies have demonstrated the importance of S-glutathiolation of proteins under conditions of oxidative or nitrosative stress.^[28,29] To prevent irreversible loss of intracellular and extracellular protein function under such stress, mixed disulfides are formed between protein cysteines and GSH. These S-glutathiolated proteins are more stable and can be dethiolated by either non-enzymatic reduction or enzymatic cleavage of the disulfide bond. Thus, S-glutathiolation allows for reversible regulation, and therefore generalized protection, of sensitive proteins.

A disrupted systemic antioxidant shield leads to predictable damage to lung tissue by oxidants. Oxidants directly harm sensitive lung epithelia. In addition, they are able to inactivate antiproteases, which then leads to increased elastase damage, increased mucus secretion, and deranged immune signaling.^[30-34] Oxidants also adversely affect ciliary beat function in the lung, and lung surfactant levels are diminished by the oxidant burden.^[33] There is an increased production of chloramines, which further decreases epithelial integrity.^[35-37] A higher oxidant burden also creates cell structure abnormalities, which may lead to impaired cell function or even premature cell death.^[38-53] In the lung, generalized bronchoconstriction can be another consequence of decreased antioxidant functioning.^[54] Damage to the epithelial tissue of the lung also permits greater adhesion of pathogens.^[55] Oxidants can also inactivate other parts of the GSH system, such as GSH reductase and γ -glutamylcysteine transferase, both necessary for cellular protection and proper redox functionality.^[56-58] Protein S-glutathiolation will decrease, resulting in irreversible loss of sensitive protein function.^[29]

A second consequence of impaired GSH efflux is increased viscosity of mucus. GSH plays an important role in mucolysis of disulfide bonds in mucus, in much the same manner as the more well known cysteine donors such as N-acetylcysteine (NAC).^[59] Increased viscosity of mucus has important consequences to the lung environment.^[59-67]

Finally, the redox system of GSH, as indicated by the GSH : GSSG ratio manifesting redox potential, is an important immune system signal. The GSH : GSSG ratio is usually greater than 9 : 1, sometimes reaching over 100–200 : 1, depending on the compartment. When that ratio is substantially decreased or there is a decrease in total GSH (GSH + GSSG), the body appears to interpret such events as a call for assistance from the immune system to cope with some threatening challenge that is resulting in pathologic oxidative reactions that are outpacing GSH replenishment. For example, Day and colleagues^[68,69] have found that when normal mouse lung tissue is challenged with *Pseudomonas aeruginosa*, there is a 3-fold induction in epithelial lining fluid (ELF) GSH levels and a 2-fold induction in CFTR levels, presumably to offset increased oxidative reactions. The inability to effect this large increase in ELF GSH because of ineffectual transport due to CFTR mutation will substantially alter both the redox ratio and the level of total GSH.

It is important to understand how the body reads the effects of CFTR absence or malfunction on the GSH system. Cells without channels redundant to CFTR, such as lung epithelia (whose channels redundant to the CFTR are at the basolateral, not the apical, surface), will not be able to export GSH to the extracellular milieu, and the extracellular deficit may become quite severe. Levels of

total GSH within such cells may remain normal, but the GSH : GSSG ratio may become substantially decreased. However, immune system cells are among the class of cells that have channels redundant to the CFTR. With a growing extracellular deficit, immune system cells may actually attempt to efflux GSH to rectify that deficit. Furthermore, with the increasing oxidant burden, immune system cells may use up their stores of GSH in self-protection. Immune cells then become GSH deficient. All in all, what the body senses in cystic fibrosis is that there is some threat that is using up all of the GSH in oxidative reactions, even though that threat is nonexistent. What is really occurring in cystic fibrosis is defective GSH efflux, but the body has no way of telling the difference.

The body responds by mobilizing itself to meet the nonexistent threat. In short, it inflames. GSH deficiency in leukocytes causes increased release of oxidants such as hydrogen peroxide.^[70] Cellular GSH deficiency causes increased transcription of nuclear factor- κ B, which then codes for greater levels of inflammatory cytokines, such as tumor necrosis factor- α , activator protein-1, monocyte chemoattractant protein-1, IL-8, and IL-1 α .^[71-85] Such a cytokine profile creates inflammation and recruitment of neutrophils and macrophages even in the absence of a threat, which is precisely what occurs in cystic fibrosis. (Of course, when a pathogen threat does present itself, the inflammation becomes even more excessive.) As long as full GSH replenishment cannot occur (because of defective GSH efflux from most of the cells of the body), the inflammation will continue and become chronic, as it is in cystic fibrosis.

In addition to chronic inflammation, the continuing inability to replenish GSH, especially in immune cells, creates a situation of immune incompetence. GSH deficiency in leukocytes causes, in general, impaired release of lysosomal enzymes, decreased phagocytosis, and premature apoptosis.^[70,86-100] GSH deficiency also creates a situation of incomplete immune system signaling, because GSH reduction of disulfide bonds is necessary for such signaling. For example, antigen-presenting cells use the reductive power of GSH to present antigens to T cells.^[101-104] B cells appear similarly affected,^[105,106] and activation of T and B cells appears related to GSH levels.^[107-112] Interferon- γ signaling is also dependent on the presence of GSH.^[113] Such interruptions of appropriate immune signaling begin to shift the organism to a more T helper-2 type of response, which is less effective in pathogen clearing.^[101,107,114-118] In addition, GSH is necessary to create a reservoir of nitric oxide (NO) [via creation of s-nitrosoglutathione (GSNO)], and the lack of such a reservoir leads to a generalized lack of NO itself in the lung environment.^[119-130] NO not only has important bactericidal properties but also is necessary in cell signaling and smooth muscle relaxation and helps regulate ciliary

beat function.^[131-137] When antioxidant defenses are compromised, superoxide anion will scavenge NO almost instantaneously, the reaction limited only by the extent of diffusion. The depletion of NO has important consequences for both lung function and immune function.

In short, then, a generalized GSH deficiency will cause inflammation coupled, paradoxically, with decreased immune system competence to clear pathogens. These effects are in addition to the loss of antioxidant protection and mucolytic activity noted above in connection with GSH deficiency. The CFTR mutations that cause cystic fibrosis produce these consequences as a result of severely impaired efflux of GSH from most cells of the body. Figure 1 represents a summary of how diminished GSH transport influences the pathophysiology of cystic fibrosis.

2. GSH System Dysfunction in Cystic Fibrosis

The evidence for a primary GSH system dysfunction in cystic fibrosis is steadily growing.

2.1 Effect of Cystic Fibrosis Transmembrane Regulator Mutation on Efflux of GSH

In retrospect, it was the work of Linsdell and Hanrahan^[138] that first identified that the CFTR channel played a role in the efflux of GSH. After clamping the CFTR channels of Chinese hamster ovary cells, Linsdell and Hanrahan^[138] compiled a list of substances that were subsequently not effluxed. GSH was on the list. Since that study published in 1998, however, it was the work of Gao and colleagues^[139] that arguably pushed that insight further. Using cell lines of cystic fibrosis lung epithelia, this team was able to demonstrate markedly decreased GSH efflux. Velsor and colleagues^[140] and Day and colleagues^[69] also found a 50% reduction in GSH levels in the ELF of the lung in uninfected CFTR knockout mice and a lack of normal induction of GSH in the ELF when challenged by *P. aeruginosa*. Kogan and colleagues^[141,142] found this same diminished GSH efflux with a variety of CFTR mutants, including G551D, R347D, K464A, and K1250A, and through the use of sophisticated tests were able to confirm that purified CFTR protein alone directly mediated nucleotide-dependent GSH flux and not via other associated chloride transport proteins.^[142] FINDER et al.^[143] also document a 49% reduction of GSH in bronchoalveolar lavage fluid from cystic patients with fibrosis. The findings

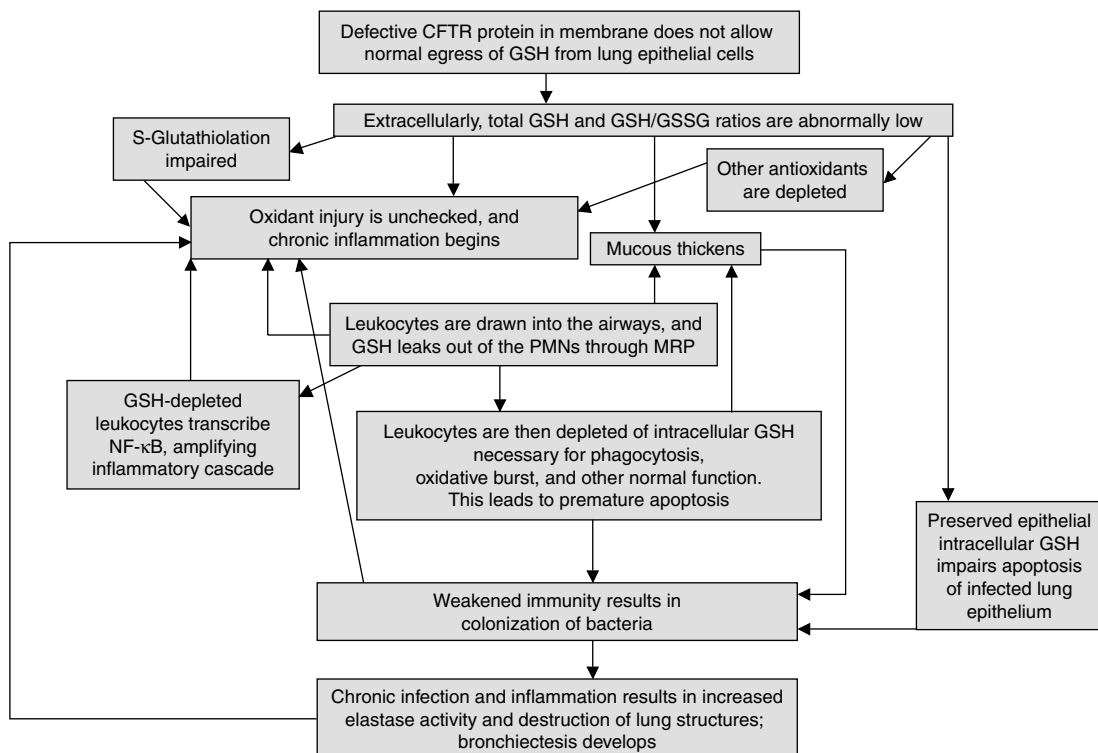


Fig. 1. The influence of diminished glutathione transport on the pathophysiology of cystic fibrosis. **CFTR** = cystic fibrosis transmembrane regulator; **GSH** = glutathione; **GSSG** = glutathione disulfide; **MRP** = multidrug resistance-associated protein; **NF-κB** = nuclear transcription factor κB; **PMN** = polymorphonuclear leukocyte.

from these five research teams constitute the 'smoking gun': cystic fibrosis directly causes significantly impaired GSH efflux.

2.2 GSH Deficiency in the Cystic Fibrosis Lung Environment

If we conceive of three distinct compartments in the cystic fibrosis lung environment, we find empirical evidence that GSH deficiency arises in the two compartments where the deficiency would be expected if a transport abnormality were at fault. The three compartments to visualize are: (i) cells that do not have a channel redundant to the CFTR (at least at the apical surface), such as the epithelium of the lung; (ii) the extracellular milieu, composed primarily of the lung ELF; and (iii) cells that do have channels redundant to the CFTR (such as leukocytes or erythrocytes).

Gao and colleagues^[139] found normal levels of total GSH in the first compartment, which included cystic fibrosis lung epithelial cells that did not have a channel redundant to the CFTR (at least at the apical surface). This should be expected, as there is no GSH synthesis defect in cystic fibrosis as there is, say, in AIDS. The GSH : GSSG ratio in these cells was not ascertained and remains to be analyzed. Note that the lack of ability to be depleted of GSH may lead to decreased levels of appropriate apoptosis when these cells are infiltrated by pathogens.

In the second compartment, consisting of ELF, several studies have found progressive GSH deficiency arising and persisting over time in patients with cystic fibrosis. Hull et al.^[144] found that non-infected cystic fibrosis do not appear to have a GSH deficiency in their ELF, though only the total GSH, and not the GSH : GSSG ratio, was analyzed. An altered GSH : GSSG ratio in the ELF of infants with cystic fibrosis would be evidence that the impaired GSH efflux has begun to have an impact in the respiratory system of these patients.^[145,146] Hull et al.^[144] did find that infected infants with cystic fibrosis infants had slightly lower total levels of GSH in their ELF. Brown et al.^[147] determined that, beyond infancy, plasma sulfhydryls decreased significantly with age in patients with cystic fibrosis. Roum et al.^[148] found a profound deficiency of GSH in the ELF in adult patients with cystic fibrosis, with levels of 5–10% of normal when oxidant burden was factored in. This team also found plasma GSH levels of about 50% of normal in these adult patients with cystic fibrosis; in both the ELF and plasma, they also found an extremely decreased GSH : GSSG ratio. Finder et al.^[143] found a 49% reduction of GSH in bronchoalveolar lavage fluid from patients with cystic fibrosis.

In the third compartment, including cells with channels redundant to the CFTR, in the lung environment, Tirouvanziam et al.^[149]

found substantially lower levels of GSH in sputum neutrophils from patients with cystic fibrosis compared with blood neutrophils, which also correlated with cell death rates. Furthermore, in this compartment, alterations in the expression of channels redundant to the CFTR have also been noted: for example, multidrug resistance-associated protein 1 has been shown to be upregulated in cystic fibrosis erythrocytes.^[150]

2.3 Related Phenomena

As part of this hypothesis of mutation-derived GSH efflux impairment, we should find that, other than the effects of the mutation, the rest of the GSH antioxidant system remains intact in patients with cystic fibrosis. This is in fact the case: at least normal levels of GSH peroxidase and γ -glutamylcysteine synthetase and increased levels of γ -glutamylcysteine transferase, γ -glutamyl transpeptidase, and GSH reductase have been found in patients with cystic fibrosis.^[17,144,148,151-153] Therefore, the observed GSH deficiency is a primary deficiency not caused by malfunctions in other parts of the overall GSH antioxidant system.

2.4 Impaired GSH Efflux and Cystic Fibrosis Pathology

Many of the effects that would be expected from impaired GSH efflux in most cells have been noted as part of cystic fibrosis pathology.

Inflammation in the absence of pathogen challenge has been noted in the youngest of infants with cystic fibrosis.^[1,2,154-157] Other antioxidant systems of the body have been found to be compromised in cystic fibrosis. This is in part a result of fat malabsorption due to pancreatic insufficiency in most patients with cystic fibrosis, resulting in lower levels of retinol and tocopherol, but this situation can be accelerated and aggravated by GSH deficiency. Affected systems include at least retinol, beta-carotene, tocopherol, activity of GSH peroxidase, ascorbic acid, and activity of superoxide dismutase.^[17,158-163] Antiproteases have been shown to be neutralized in cystic fibrosis, and surfactant levels are lower.^[164,165] There is an altered cytokine profile consonant with GSH deficiency, and also exhaled NO is not elevated, as it is in other respiratory diseases with a high oxidant burden.^[1-8,166-178] Interestingly, *in vitro*, the addition of S-nitrosoglutathione to delF508 cystic fibrosis cell lines has been shown to help in the maturation and functionality of the mutated protein.^[179,180] Furthermore, decreased apoptosis has been noted in pathogen-infiltrated cells without redundant anion channels in cystic fibrosis.^[181]

In conclusion, the view that CFTR acts as an important GSH efflux channel is gaining strength through recent empirical research findings. In addition to several 'smoking guns', the related

phenomena and effects that one would expect if cystic fibrosis caused GSH efflux impairment are also empirically demonstrable (figure 1). This evidence leads us to inquire about therapeutic implications. However, it is important to remember that cystic fibrosis pathophysiology is multifactorial, and rectification of GSH system dysfunction in patients with cystic fibrosis will not eliminate all harmful effects of the disease.

3. Therapeutic Implications

The usual and most direct route to augment GSH levels is to provide a cysteine donor, such as NAC, to the patient. As cysteine is the rate-limiting amino acid for GSH synthesis, this route is generally effective in otherwise healthy individuals. However, as we have seen, GSH synthesis is not impaired in cystic fibrosis; the problem is in GSH efflux from the cells in which GSH is synthesized. Nevertheless, Hosseini and colleagues^[182] have used a cysteine-rich whey powder to treat C57B1/6 mice infected with *Pseudomonas* spp. and noted some improvement in mortality; therefore, precursors might usefully complement a strategy of exogenous GSH augmentation.

Direct augmentation of GSH levels in the ELF with aerosolized GSH has been carried out *in vivo*, including in patients with cystic fibrosis, AIDS, idiopathic pulmonary fibrosis (IPF), COPD, and other diseases.^[183-191] Unfortunately, GSH in solution has a pH of 2.7 and is an irritant to the lung. This has hampered the usefulness of this therapy for patients with respiratory ailments. Two clinical trials, using buffered GSH with a pH of 5–6, have been carried out in patients with cystic fibrosis.

In the first trial, using the AKITA®¹ inhalation device, Griese et al.^[192] were able to increase the GSH level in bronchoalveolar lavage fluid in patients with cystic fibrosis through inhalation of a buffered GSH solution. One hour after inhalation, GSH levels increased 3- to 4-fold, and at 12 hours levels of GSH were still almost double those at baseline. Griese et al. found that with 14 days' use of three times daily buffered GSH 300–450mg, FEV₁ and FVC increased an average of 6–7% over baseline ($p < 0.001$).^[193] No change in oxidative markers was observed, though this might be because of the short duration of therapy.

The second study carried out by Bishop et al.^[194,195] was a randomized, double-blind, placebo-controlled trial. The dosage of buffered GSH was 66 mg/kg/day, divided into four inhalation sessions over a 6-week period. Results indicated that 11 of 13 clinical indicators examined favored the GSH treatment group over the placebo group, including lung function scores, and statistical significance was achieved in improvement in several of the

indicators, including peak flow, and in compliance analysis, cough.

The results of these two clinical trials of aerosolized buffered GSH are very promising and warrant larger, multicenter trials of longer duration.

Oral administration of GSH is not to be overlooked. Previously, researchers could not agree on whether GSH was cleaved in the digestive tract or taken up intact in the jejunum. Newer studies seem to indicate the latter.^[196-200] Furthermore, there is new and innovative research being conducted to create a novel peptide that could serve as a GSH efflux for cystic fibrosis cells.^[201] Finally, intravenous GSH has been used as a treatment for radiation poisoning, as well as for other diseases such as Parkinson disease.^[183,202,203] Given that it is most likely that the lung is a net importer of circulating GSH, this route might bear further investigation in the case of cystic fibrosis.

Other respiratory ailments are marked by a decrease in GSH in the ELF. Clinicians treating such illnesses may want to examine GSH augmentation in diseases such as adult respiratory distress syndrome (ARDS), COPD, idiopathic interstitial pneumonia, IPF of nonsmokers, idiopathic respiratory distress syndrome, and diffuse fibrosing alveolitis.

In summary, augmentation of GSH in the ELF is feasible and may be useful not only for cystic fibrosis, but also for several other respiratory conditions. Given the vasodilatory and anti-inflammatory properties of GSH, there may be contraindications to its use. It could be speculated that in patients with a history of hemoptysis/pneumothorax, those yielding a positive culture for *Burkholderia cepacia*, or those with an FEV₁ <30% predicted, the use of GSH may be contraindicated until further, more extensive trials have been conducted.

4. Conclusion

In conclusion, new research is beginning to alter our understanding of the CFTR channel. It is no longer possible to view it merely or even primarily as a chloride efflux channel. At this point in time, it must be viewed as a chloride/GSH/bicarbonate channel, though this list may grow in the future. As we more fully understand the nature and functions of the CFTR channel, new therapeutic approaches will begin to come into view, as we have seen with GSH. In one respect, cystic fibrosis may be viewed (at least in part) as the first identified disease with GSH transport dysfunction.

Clinicians may be able to make effective use of these new insights from cutting-edge research. However, it is important to remember that the pathophysiology of cystic fibrosis is multifactorial, and rectification of GSH system dysfunction in cystic fibrosis

1 The use of trade names is for product identification purposes only and does not imply endorsement.

patients will not eliminate all harmful effects of the disease. Indeed, some clinicians report that a significant proportion of their patients with cystic fibrosis are already using GSH and/or NAC without the physician's knowledge.^[204] Clinicians should inquire from their patients with cystic fibrosis if they are already using GSH/NAC, in order to monitor subsequent developments. In addition, patients may be unaware of possible contraindications, the importance of pH adjustment for aerosol use, or the inherent problems connected with NAC or other cysteine donor use alone. In short, given increasing patient interest in a growing body of empirical evidence demonstrating mutation-derived GSH system dysfunction in cystic fibrosis, it behooves the clinician to learn as much about this area of research as possible in order to treat cystic fibrosis patients in a more informed manner. Furthermore, clinicians may find GSH augmentation a useful intervention for other respiratory diseases, as noted in section 3, such as COPD, ARDS, and IPF.

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References

- Khan TZ, Wagener JS, Bost T, et al. Early pulmonary inflammation in infants with cystic fibrosis. *Am J Respir Crit Care Med* 1995 Apr; 151 (4): 1075-82
- Balough K, McCubbin M, Weinberger M, et al. The relationship between infection and inflammation in the early stages of lung disease from cystic fibrosis. *Pediatr Pulmonol* 1995 Aug; 20 (2): 63-70
- Muhlebach MS, Stewart PW, Leigh MW, et al. Quantitation of inflammatory responses to bacteria in young cystic fibrosis and control patients. *Am J Respir Crit Care Med* 1999 Jul; 160 (1): 186-91
- Noah TL, Black HR, Cheng PW, et al. Nasal and bronchoalveolar lavage fluid cytokines in early cystic fibrosis. *J Infect Dis* 1997 Mar; 175 (3): 638-47
- Osika E, Cavaillon JM, Chadelat K, et al. Distinct sputum cytokine profiles in cystic fibrosis and other chronic inflammatory airway diseases. *Eur Respir J* 1999 Aug; 14 (2): 339-46
- Tabary O, Escotte S, Couetil JP, et al. Genistein inhibits constitutive and inducible NFkappaB activation and decreases IL-8 production by human cystic fibrosis bronchial gland cells. *Am J Pathol* 1999 Aug; 155 (2): 473-81
- Dai Y, Dean TP, Church MK, et al. Desensitisation of neutrophil responses by systemic interleukin 8 in cystic fibrosis. *Thorax* 1994 Sep; 49 (9): 867-71
- Bonfield TL, Konstan MW, Berger M. Altered respiratory epithelial cell cytokine production in cystic fibrosis. *J Allergy Clin Immunol* 1999; 104 (1): 72-8
- Pilewski JM, Frizzell RA. Role of CFTR in airway disease. *Physiol Rev* 1999 Jan; 79 Suppl. 1: S215-55
- van der Vliet A, Eiserich JP, Marelich GP, et al. Oxidative stress in cystic fibrosis: does it occur and does it matter? *Adv Pharmacol* 1997; 38: 491-513
- Tamada T, Hug M, Peters K, et al. Bicarbonate secretion in airway serous cells. *Pediatr Pulmonol* 2001 Oct; 22: 119-20
- Coakley RD. Regulation of airway surface liquid pH in cystic fibrosis. *Pediatr Pulmonol* 2001 Oct; 22: 120-1
- McShane D, Davies JC, Davies MG, et al. Airway surface pH in subjects with cystic fibrosis. *Eur Respir J* 2003; 21 (1): 37-42
- Jayaraman S, Joo NS, Reitz B, et al. Submucosal gland secretions in airways from cystic fibrosis patients have normal [NA(+)] and pH but elevated viscosity. *Proc Natl Acad Sci U S A* 2001; 98 (14): 8119-23
- Jayaraman S, Song Y, Verkman AS. Airway surface liquid pH regulation in well-differentiated airway epithelial cell cultures and mouse trachea. *Am J Physiol Cell Physiol* 2001; 281 (5): C1504-11
- Hudson VM. Rethinking cystic fibrosis pathology: the critical role of abnormal reduced glutathione (GSH) transport caused by CFTR mutation. *Free Radic Biol Med* 2001; 30 (12): 1440-61
- Benabdeslam H, Abidi H, Garcia I, et al. Lipid peroxidation and antioxidant defenses in cystic fibrosis patients. *Clin Chem Lab Med* 1999 May; 37 (5): 511-6
- Stocker R, Bowry VW, Frei B, et al. Ubiquinol-10 protects human low density lipoprotein more efficiently against lipid peroxidation than does alpha-tocopherol. *Proc Natl Acad Sci U S A* 1991 Mar; 88: 1646-50
- Thomas SR, Neuzil J, Stocker R. Cosupplementation with coenzyme Q prevents the prooxidant effect of alpha-tocopherol and increases the resistance of LDL to transition metal-dependent oxidation initiation. *Atheroscler Thromb Vasc Biol* 1996 May; 16 (5): 687-96
- Thanissarl J, Raveendran M, Devaraj H. Buthionine sulfoximine-induced glutathione depletion: its effect on antioxidants, lipid peroxidation and calcium homeostasis in the lung. *Biochem Pharmacol* 1995 Jul 17; 50 (2): 229-34
- Leedle RA, Aust SD. The effect of glutathione on the vitamin E requirement for inhibition of liver microsomal lipid peroxidation. *Lipids* 1990 May; 25 (5): 241-5
- Higuchi Y, Matsukawa S. Glutathione depletion induces giant DNA and high-molecular-weight DNA fragmentation associated with apoptosis through lipid peroxidation and protein kinase C activation in C6 glioma cells. *Arch Biochem Biophys* 1999 Mar 1; 363 (1): 33-42
- Scholz RW, Reddy PV, Wynn MK, et al. Glutathione-dependent factors and inhibition of rat liver microsomal lipid peroxidation. *Free Radic Biol Med* 1997; 23 (5): 815-28
- Hagiwara K, Naito K, Kurokawa Y, et al. Kidney injury induced by lipid peroxide produced by vitamin E deficiency and GSH depletion in rats. *J Nutr Sci Vitaminol (Tokyo)* 1991 Feb; 37 (1): 99-107
- Angelini P, Cirelli F, Quarticelli A, et al. Administration of glutathione and lipid peroxidation induced during fasting. *Boll Soc Ital Biol Sper* 1990 Nov; 66 (11): 1097-104
- Rosenblat M, Aviram M. Macrophage glutathione content and glutathione peroxidase activity are inversely related to cell-mediated oxidation of LDL: in vitro and in vivo studies. *Free Radic Biol Med* 1998 Jan 15; 24 (2): 305-17
- Chen Q, Galleano M, Cederbaum AI. Cytotoxicity and apoptosis produced by arachidonic acid in Hep G2 cells overexpressing human cytochrome P4502E1. *J Biol Chem* 1997 Jun 6; 272 (23): 14532-41
- Klatt P, Lamas S. Regulation of protein function by S-glutathiolation in response to oxidative and nitrosative stress. *Eur J Biochem* 2000 Aug; 267 (16): 4928-44
- Thomas JA, Mallis RJ. Aging and oxidation of reactive protein sulfhydryls. *Exp Gerontol* 2001 Sep; 36 (9): 1519-26
- Buhl R, Meyer A, Vogelmeier C. Oxidant-protease interaction in the lung: prospects for antioxidant therapy. *Chest* 1996 Dec; 110 (6 Suppl.): 267S-72S
- Gillissen A, Birrer P, McElvaney NG, et al. Recombinant secretory leukoprotease inhibitor augments glutathione levels in lung epithelial lining fluid. *J Appl Physiol* 1993 Aug; 75 (2): 825-32
- Vogelmeier C, Biedermann T, Maier K, et al. Comparative loss of activity of recombinant secretory leukoprotease inhibitor and alpha 1-protease inhibitor caused by different forms of oxidative stress. *Eur Respir J* 1997 Sep; 10 (9): 2114-9
- Barbero GJ. Therapeutic approaches to cystic fibrosis. *Bull World Health Organ* 1994; 72 (3): 341-52
- Brenneisen P, Briviba K, Wlaschek M, et al. Hydrogen peroxide (H2O2) increases the steady-state mRNA levels of collagenase/MMP-1 in human dermal fibroblasts. *Free Radic Biol Med* 1997; 22 (3): 515-24
- Ogino T, Packer L, Maguire JJ. Neutrophil antioxidant capacity during the respiratory burst: loss of glutathione induced by chloramines. *Free Radic Biol Med* 1997; 23 (3): 445-52
- Bilzer M, Lauterberg BH. Glutathione metabolism in activated human neutrophils: stimulation of glutathione synthesis and consumption of glutathione by reactive oxygen species. *Eur J Clin Invest* 1991 Jun; 21 (3): 316-22

37. Vanglarik CJ, Giron-Calle J, Matalon S, et al. Extracellular glutathione protects airway epithelial cells against oxidative damage caused by hypochlorous acid exposure. *Pediatr Pulmonol* 2001 Oct; 22: 276-7
38. Freeman ML, Huntley SA, Meredith MJ, et al. Destabilization and denaturation of cellular protein by glutathione depletion. *Cell Stress Chaperones* 1997 Sep; 2 (3): 191-8
39. Calabreses V, Testa G, Ravagna A, et al. HSP70 induction in the brain following ethanol administration in the rat: regulation by glutathione redox state. *Biochem Biophys Res Commun* 2000 Mar 16; 269 (2): 397-400
40. Liu H, Lightfoot R, Stevens JL. Activation of heat shock factor by alkylating agents is triggered by glutathione depletion and oxidation of protein thiols. *J Biol Chem* 1996 Mar 1; 271 (9): 4805-12
41. Wilhelm D, Bender K, Knebel A, et al. The level of intracellular glutathione is a key regulator for the induction of stress-activated signal transduction pathways including Jun N-terminal protein kinases and p38 kinase by alkylating agents. *Mol Cell Biol* 1997 Aug; 17 (8): 4792-800
42. Kirilin WG, Cai J, Thompson SA, et al. Glutathione redox potential in response to differentiation and enzyme inducers. *Free Radic Biol Med* 1999 Dec; 27 (11-12): 1208-18
43. Lertratanangkoon K, Savaraj N, Scimeca JM, et al. Glutathione depletion-induced thymidylate insufficiency for DNA repair synthesis. *Biochem Biophys Res Commun* 1997 May 19; 234 (2): 470-5
44. Asensi M, Garcia-Espana A, Pallardo FV, et al. Effect of nonprotein thiols on protein synthesis in isolated rat hepatocytes. *Experientia* 1996 Feb 15; 52 (2): 111-4
45. Fussell JC, Kelly FJ. Effects of dexamethasone on lung protein turnover. *Biochem J* 1991 Jan 1; 273 (Pt 1): 93-7
46. Murthy MR. Protein synthesis in growing-rat tissues: II. Polyribosome concentration of brain and liver as a function of age. *Biochim Biophys Acta* 1966 Jun 22; 119 (3): 599-613
47. Thanissar J, Raveendran M, Sivasithamparan N, et al. Effect of chronic glutathione deficiency on rat lung mitochondrial function. *Pulm Pharmacol* 1996 Apr; 9 (2): 119-22
48. Rojas E, Valverde M, Kala SV, et al. Accumulation of DNA damage in the organs of mice deficient in gamma-glutamyltranspeptidase. *Mutat Res* 2000 Feb 14; 447 (2): 305-16
49. Gilmont RR, Dardano A, Young M, et al. Effects of glutathione depletion on oxidant-induced endothelial cell injury. *J Surg Res* 1998 Nov; 80 (1): 62-8
50. Fiorentini C, Falzano L, Rivabene R, et al. N-acetylcysteine protects epithelial cells against the antioxidant imbalance due to *Clostridium difficile* toxins. *FEBS Lett* 1999 Jun 18; 453 (1-2): 124-8
51. Kamei H. Cystine starvation induces reversible large-body formation from nuclear bodies in T24 cells. *Exp Cell Res* 1997 Nov 25; 237 (1): 207-16
52. Zuelke KA, Jones DP, Perreault SD. Glutathione oxidation is associated with altered microtubule function and disrupted fertilization in mature hamster oocytes. *Biol Reprod* 1997 Dec; 57 (6): 1413-9
53. Jain A, Martensson J, Mehta T, et al. Ascorbic acid prevents oxidative stress in glutathione deficient mice: effects on lung type 2 cell lamellar bodies, lung surfactant, and skeletal muscle. *Proc Natl Acad Sci U S A* 1992 Jun; 89 (11): 5093-7
54. Shaheen SO, Sterne JA, Songhurst CE, et al. Frequent paracetamol use and asthma in adults. *Thorax* 2000 Apr; 55 (4): 266-70
55. Li XY, Donaldson K, Rahman I, et al. An investigation of the role of glutathione in increased epithelial permeability induced by cigarette smoke in vivo and in vitro. *Am J Respir Crit Care Med* 1994 Jun; 149 (6): 1518-25
56. Remiao F, Carmo H, Carvalho FD, et al. Inhibition of glutathione reductase by isoproterenol oxidation products. *J Enzyme Inhib* 2000; 15 (1): 47-61
57. Barker JE, Heales SJ, Cassidy A, et al. Depletion of brain glutathione results in a decrease of glutathione reductase activity: an enzyme susceptible to oxidative damage. *Brain Res* 1996 Apr 15; 16 (1-2): 118-22
58. van Klaveren RJ, Hoet PH, Pype JL, et al. Increase in gamma-glutamyltransferase by glutathione depletion in rat type II pneumocytes. *Free Radic Biol Med* 1997; 22 (3): 525-34
59. Smith CV, Jones DP, Guenther TM, et al. Compartmentation of glutathione: implications for the study of toxicity and disease. *Toxicol Appl Pharmacol* 1996; 140: 1-12
60. Houtmeyers E, Gosselink R, Gayan-Ramirez G, et al. Regulation of mucociliary clearance in health and disease. *Eur Respir J* 1999 May; 13 (5): 1177-88
61. King M, Rubin BK. Mucus-controlling agents: past and present. *Respir Care Clin N Am* 1999 Dec; 5 (4): 575-94
62. Houtmeyers E, Gosselink R, Gayan-Ramirez G, et al. Effects of drugs on mucus clearance. *Eur Respir J* 1999 Aug; 14 (2): 452-67
63. Connolly MA. Mucolytics and the critically ill patient: help or hindrance? *AACN Clin Issues* 1995 May; 6 (2): 307-15
64. Clarke SW. Rationale of airway clearance. *Eur Respir J Suppl* 1989 Jul; 7: 599s-603s
65. Jiang N, Dreher KL, Dye JA, et al. Residual oil fly ash induces cytotoxicity and mucin secretion by guinea pig tracheal epithelial cells via an oxidant-mediated mechanism. *Toxicol Appl Pharmacol* 2000 Mar 15; 163 (3): 221-30
66. Li JD, Feng W, Gallup M, et al. Activation of NF-kappaB via a Src-dependent Ras-MAPK-pp90rsk pathway is required for *Pseudomonas aeruginosa*-induced mucin overproduction in epithelial cells. *Proc Natl Acad Sci U S A* 1998; 95 (10): 5718-23
67. Fischer B, Voynow J. Neutrophil elastase induces MUC5AC messenger RNA expression by an oxidant-dependent mechanism. *Chest* 2000 May; 117 (5 Suppl. 1): 317S-20S
68. Day BJ, van Heeckeren A, Velsor LW. Elevation of lung CFTR, MRP-2, and epithelial lining fluid glutathione in mice with *Pseudomonas aeruginosa* lung infection [abstract]. *Am J Respir Crit Care Med* 2003 Apr; 167 (7): A916
69. Day BJ, van Heeckeren AM, Min E, et al. Role for cystic fibrosis transmembrane conductance regulator protein in a glutathione response to bronchopulmonary pseudomonas infection. *Infect Immun* 2004 Apr; 72 (4): 2045-51
70. Boxer LA, Oliver JM, Speilberg SP, et al. Protection of granulocytes by vitamin E in glutathione synthetase deficiency. *N Engl J Med* 1979 Oct 25; 301 (17): 901-5
71. Cho S, Urata Y, Iida T, et al. Glutathione downregulates the phosphorylation of I kappa-B: autoloop regulation of the NF-kappa B-mediated expression of NF-kappa B subunits by TNF-alpha in mouse vascular endothelial cells. *Biochem Biophys Res Commun* 1998; 253 (1): 104-8
72. Droge W, Eck HP, Gmunder H, et al. Modulation of lymphocyte functions and immune responses by cysteine and cysteine derivatives. *Am J Med* 1991 Sep 30; 91 (3C): 140-3
73. Sen CK, Khanna S, Reznick AZ, et al. Glutathione regulation of tumor necrosis factor-alpha-induced NF-kappa B activation in skeletal muscle-derived L6 cells. *Biochem Biophys Res Commun* 1997 Aug 28; 237 (3): 645-9
74. Desai A, Huang X, Warren JS. Intracellular glutathione redox status modulates MCP-1 expression in pulmonary granulomatous vasculitis. *Lab Invest* 1999 Jul; 79 (7): 837-47
75. Luo Y, Hattori A, Munoz J, et al. Intraatrial dopamine injection induces apoptosis through oxidation-involved activation of transcription factors AP-1 and NF-kappaB in rats. *Mol Pharmacol* 1999 Aug; 56 (2): 254-64
76. Haddad JJ, Olver RE, Land SC. Antioxidant/pro-oxidant equilibrium regulates HIF-1alpha and NF-kappaB redox sensitivity: evidence for inhibition by glutathione oxidation in alveolar epithelial cells. *J Biol Chem* 2000; 275 (28): 21130-9
77. Tanaka C, Kamata H, Takeshita H, et al. Redox regulation of lipopolysaccharide (LPS)-induced interleukin-8 (IL-8) gene expression mediated by NF kappa B and AP-1 in human astrocytoma U373 cells. *Biochem Biophys Res Commun* 1997 Mar 17; 232 (2): 568-73
78. Rahman I, MacNee W. Regulation of redox glutathione levels and gene transcription in lung inflammation: therapeutic approaches. *Free Radic Biol Med* 2000 May 1; 28 (9): 1405-20
79. Christman JW, Sadikot RT, Blackwell TS. The role of nuclear factor-kappa B in pulmonary diseases. *Chest* 2000; 117 (5): 1482-7
80. Gosset P, Wallaert B, Tonnel AB, et al. Thiol regulation of the production of TNF-alpha, IL-6 and IL-8 by human alveolar macrophages. *Eur Respir J* 1999 Jul; 14 (1): 98-105
81. Hashimoto S, Gon Y, Matsumoto K, et al. Regulation by intracellular glutathione of TNF-alpha-induced p38 MAP kinase activation and RANTES production by human pulmonary vascular endothelial cells. *Allergy* 2000 May; 55 (5): 463-9
82. Yamauchi N, Watanabe N, Kuriyama H, et al. Suppressive effects of intracellular glutathione on hydroxyl radical production induced by tumor necrosis factor. *Int J Cancer* 1990 Nov 15; 46 (5): 884-8

83. Peristeris P, Clark BD, Gatti S, et al. N-acetylcysteine and glutathione as inhibitors of tumor necrosis factor production. *Cell Immunol* 1992 Apr; 140 (2): 390-9
84. Rovin BH, Dickerson JA, Tan LC, et al. Modulation of IL-1-induced chemokine expression in human mesangial cells through alterations in redox status. *Cytokine* 1997 Mar; 9 (3): 178-86
85. Pena LR, Hill DB, McClain CJ. Treatment with glutathione precursor decreases cytokine activity. *JPEN J Parenter Enteral Nutr* 1999 Jan-Feb; 23 (1): 1-6
86. Watson RW, Rotstein OD, Nathens AB, et al. Thiol-mediated redox regulation of neutrophil apoptosis. *Surgery* 1996 Aug; 120 (2): 150-7
87. Nakatani T, Tawaramoto M, Opare Kennedy D, et al. Apoptosis induced by chelation of intracellular zinc is associated with depletion of cellular reduced glutathione level in rat hepatocytes. *Chem Biol Interact* 2000 Mar 15; 125 (3): 151-63
88. Coppola S, Ghibelli L. GSH extrusion and the mitochondrial pathway of apoptotic signalling. *Biochem Soc Trans* 2000 Feb; 28 (2): 56-61
89. Colell A, Garcia-Ruiz C, Miranda M, et al. Selective glutathione depletion of mitochondria by ethanol sensitizes hepatocytes to tumor necrosis factor. *Gastroenterology* 1998 Dec; 115 (6): 1541-51
90. Nicole A, Santiard-Baron D, Ceballos-Picot I. Direct evidence for glutathione as mediator of apoptosis in neuronal cells. *Biomed Pharmacother* 1998; 52 (9): 349-55
91. Rosenfeld ME. Inflammation, lipids, and free radicals: lessons learned from the atherogenic process. *Semin Reprod Endocrinol* 1998; 16 (4): 249-61
92. Hardwick SJ, Carpenter KL, Allen EA, et al. Glutathione (GSH) and the toxicity of oxidised low-density lipoprotein to human monocyte-macrophages. *Free Radic Res* 1999 Jan; 30 (1): 11-9
93. Aoshiba K, Yasui S, Nishimura K, et al. Thiol depletion induces apoptosis in cultured lung fibroblasts. *Am J Respir Cell Mol Biol* 1999 Jul; 21 (1): 54-64
94. Vahrmeijer AL, van Dierendonck JH, Schutrups J, et al. Effect of glutathione depletion on inhibition of cell cycle progression and induction of apoptosis by melphalan (L-phenylalanine mustard) in human colorectal cancer cells. *Biochem Pharmacol* 1999 Aug 15; 58 (4): 655-64
95. Yang CF, Shen HM, Ong CN. Ebselen induces apoptosis in HepG (2) cells through rapid depletion of intracellular thiols. *Arch Biochem Biophys* 2000 Feb 15; 374 (2): 142-52
96. Li Y, Maher P, Schubert D. A role for 12-lipoxygenase in nerve cell death caused by glutathione depletion. *Neuron* 1997 Aug; 19 (2): 453-63
97. Iwata S, Hori T, Sato N, et al. Adult T cell leukemia (ATL)-derived factor/human thioredoxin prevents apoptosis of lymphoid cells induced by L-cystine and glutathione depletion: possible involvement of thiol-mediated redox regulation in apoptosis caused by pro-oxidant state. *J Immunol* 1997 Apr; 158 (7): 3108-17
98. Wedner HJ, Simchowicz L, Stenson WF, et al. Inhibition of human polymorphonuclear leukocyte function by 2-cyclohexene-1-one: A role for glutathione in cell activation. *J Clin Invest* 1981 Aug; 68 (2): 535-43
99. Aziz AS, Klesius PH, Frandsen JC. Effects of selenium on polymorphonuclear leukocyte function in goats. *Am J Vet Res* 1984 Sep; 45 (9): 1715-8
100. Voetman AA, Loos JA, Roos D. Changes in the levels of glutathione in phagocytosing human neutrophils. *Blood* 1980 May; 55 (5): 741-7
101. Peterson JD, Herzenberg LA, Vasquez K, et al. Glutathione levels in antigen-presenting cells modulate Th1 versus Th2 response patterns. *Proc Natl Acad Sci U S A* 1998 Mar 17; 95 (6): 3071-6
102. Nathens AB, Rotstein OD, Dackiw AP, et al. The glutathione depleting agent diethylmaleate prolong renal allograft survival. *J Surg Res* 1998 Jun; 77 (1): 75-9
103. Robinson MK, Rodrick ML, Jacobs DO, et al. Glutathione depletion in rats impairs T-cell and macrophage immune function. *Arch Surg* 1993 Jan; 128 (1): 29-34
104. Ginn-Pease ME, Whisler RL. Optimal NF kappa B mediated transcriptional responses to Jurkat T cells exposed to oxidative stress are dependent on intracellular glutathione and costimulatory signals. *Biochem Biophys Res Commun* 1996 Sep 24; 226 (3): 695-702
105. Jeannin P, Delneste Y, Lecoanet-Henchoz S, et al. Thiols decrease human interleukin (IL) 4 production and IL-4-induced immunoglobulin synthesis. *J Exp Med* 1995 Dec 1; 182 (6): 1785-92
106. Rigacci S, Iantomasi T, Marraccini P, et al. Evidence for glutathione involvement in platelet-derived growth-factor-mediated signal. *Biochem J* 1997 Jun 15; 324 (Pt 3): 791-6
107. Smyth MJ. Glutathione modulates activation-dependent proliferation of human peripheral blood lymphocyte populations without regulating their activated function. *J Immunol* 1991 Mar 15; 146 (6): 1921-7
108. Romero DL, Mounho BJ, Lauer FT, et al. Depletion of glutathione by benzo (a)pyrene metabolites, ionomycin, thapsigargin, and phorbol myristate in human peripheral blood mononuclear cells. *Toxicol Appl Pharmacol* 1997 May; 144 (1): 62-9
109. Liang SM, Liang CM, Hargrove ME, et al. Regulation by glutathione of the effect of lymphokines on differentiation of primary activated lymphocytes: influence of glutathione on cytotoxic activity of CD3-AK-. *J Immunol* 1991 Mar 15; 146 (6): 1909-13
110. Gmunder H, Droge W. Differential effects of glutathione depletion on T cell subsets. *Cell Immunol* 1991 Nov; 138 (1): 229-37
111. Ting CC, Hargrove HE, Liang SM, et al. Dichotomy of glutathione regulation of the activation of resting and preactivated lymphocytes. *Cell Immunol* 1992 Jun; 142 (1): 40-53
112. Gmunder H, Eck HP, Benninghoff B, et al. Macrophages regulate intracellular glutathione levels of lymphocytes: evidence for an immunoregulatory role of cysteine. *Cell Immunol* 1990 Aug; 129 (1): 32-46
113. Galiotta LJV, Folli C, Marchetti C, et al. Modification of transepithelial ion transport in human cultured bronchial epithelial cells by interferon-gamma. *Am J Physiol Lung Cell Mol Physiol* 2000 Jun; 278 (6): L1186-94
114. van der Meide PH, de Labie MC, Botman CA, et al. Mercuric chloride down-regulates T cell interferon-gamma production in brown Norway but not in Lewis rats: role of glutathione. *Eur J Immunol* 1993 Mar; 23 (3): 675-81
115. Sprietsma JE. Zinc-controlled Th1/Th2 switch significantly determines development of disease. *Med Hypotheses* 1997 Jul; 49 (1): 1-14
116. Sprietsma JE. Cysteine, glutathione (GSH) and zinc and copper ions together are effective, natural, intracellular inhibitors of (AIDS) viruses. *Med Hypotheses* 1999 Jun; 52 (6): 529-38
117. Sprietsma JE. Modern diets and diseases: NO-zinc balance: under Th1, zinc and nitrogen monoxide (NO) collectively protect against viruses, AIDS, autoimmunity, diabetes, allergies, asthma, infectious diseases, atherosclerosis and cancer. *Med Hypotheses* 1999 Jul; 53 (1): 6-16
118. Moser C, Johansen HK, Song Z, et al. Chronic pseudomonas aeruginosa lung infection is more severe in Th2 responding BALB/c mice compared to Th1 responding C3H/HeN mice. *APMIS* 1997 Nov; 105 (11): 838-42
119. Chen G, Wang SH, Warner TD. Regulation of iNOS mRNA levels in endothelial cells by glutathione, a double-edged sword. *Free Radic Res* 2000 Mar; 32 (3): 223-34
120. Kang KW, Pak YM, Kim ND. Diethylmaleate and buthionine sulfoximine, glutathione-depleting agents, differentially inhibit expression of inducible nitric oxide synthase in endotoxemic mice. *Nitric Oxide* 1999 Jun; 3 (3): 265-71
121. Vos TA, Goor H, Tuyt L, et al. Expression of inducible nitric oxide synthase in endotoxemic rat hepatocytes is dependent on the cellular glutathione status. *Hepatology* 1999 Feb; 29 (2): 421-6
122. Wang F, Wang LY, Wright D, et al. Redox imbalance differentially inhibits lipopolysaccharide-induced macrophage activation in the mouse liver. *Infect Immun* 1999 Oct; 67 (10): 5409-16
123. Davidson CA, Kaminski PM, Wolin MS. NO elicits prolonged relaxation of bovine pulmonary arteries via endogenous peroxynitrite generation. *Am J Physiol* 1997 Aug; 273 (2 Pt 1): L437-44
124. Singh SP, Wishnok JS, Keshive M, et al. The chemistry of the S-nitrosoglutathione/glutathione system. *Proc Natl Acad Sci U S A* 1996 Dec; 93: 14428-33
125. D'Emilia DM, Lipton SA. Ratio of S-nitrosomocyst(e)ine or other thiols determines neurotoxicity in rat cerebrocortical cultures. *Neurosci Lett* 1999; 265: 103-6
126. Stefanelli C, Pignatti C, Tantani B, et al. Nitric oxide can function as either a killer molecule or an antiapoptotic effector in cardiomyocytes. *Biochim Biophys Acta* 1999 Jul 8; 1450 (3): 406-13
127. Rosenberg PA, Li Y, Ai S, et al. Intracellular redox state determines whether nitric oxide is toxic or protective to rat oligodendrocytes in culture. *J Neurochem* 1999 Aug; 73 (2): 476-84
128. Chiueh CC, Rauhala P. The redox pathway of S-nitrosoglutathione, glutathione and nitric oxide in cell to neuron communications. *Free Radic Res* 1999 Dec; 31 (6): 641-50

129. Jones KL, Bryan TW, Jinkins PA, et al. Superoxide released from neutrophils causes a reduction in nitric oxide gas. *Am J Physiol* 1998 Dec; 275 (6): L1120-6
130. Zavadnik IB, Lapshina EA, Rekawiecka K, et al. Membrane effects of nitrite-induced oxidation of human red blood cells. *Biochim Biophys Acta* 1999 Oct 15; 1421 (2): 306-16
131. Kelley TJ, Drumm ML. Inducible nitric oxide synthase expression is reduced in cystic fibrosis murine and human airway epithelial cell lines. *J Clin Invest* 1998 Sep; 102 (6): 1200-7
132. Mayer B, Pfeiffer S, Schrammel A, et al. A new pathway of nitric oxide cyclic GMP signalling involving s-nitrosoglutathione. *J Biol Chem* 1998 Feb 6; 273 (6): 3264-70
133. Belvisi MG, Ward JK, Mitchell JA, et al. Nitric oxide as a neurotransmitter in human airways. *Arch Int Pharmacodyn Ther* 1995 Jan-Feb; 329 (1): 97-110
134. Gaston B, Drazen JM, Jansen A, et al. Relaxation of human bronchial smooth muscle by S-nitrosothiols in vitro. *J Pharmacol Exp Ther* 1994 Feb; 268 (2): 978-84
135. Kamosinska B, Radomski MW, Duszyk M, et al. Nitric oxide activates chloride currents in human lung epithelial cells. *Am J Physiol* 1997 Jun; 272 (6 Pt 1): L1098-104
136. Dong YJ, Chao AC, Kouyama K, et al. Activation of CFTR chloride current by nitric oxide in human T lymphocytes. *EMBO J* 1995 Jun 15; 14 (12): 2700-7
137. Runer T, Lindberg S. Ciliostimulatory effects mediated by nitric oxide. *Acta Otolaryngol* 1999; 119 (7): 821-5
138. Linsdell P, Hanrahan JW. Glutathione permeability of CFTR. *Am J Physiol* 1998 Jul; 44 (1): C323-6
139. Gao L, Kim KJ, Yankaskas JR, et al. Abnormal glutathione transport in cystic fibrosis airway epithelia. *Am J Physiol* 1999 Jul; 277 (1): L113-8
140. Velsor LW, van Heeckeren A, Day BJ. Antioxidant imbalance in the lungs of cystic fibrosis transmembrane conductance regulator protein mutant mice. *Am J Physiol Lung Cell Mol Physiol* 2001 Jul; 281 (1): L31-8
141. Kogan I, Ramjessingh M, Kidd J, et al. Characterization of glutathione permeability through the CFTR channel pore. *Pediatr Pulmonol* 2002 Oct; 24: 189-90
142. Kogan I, Ramjessingh M, Li C, et al. CFTR directly mediates nucleotide-regulated glutathione flux. *EMBO J* 2003 May 1; 22 (9): 1981-9
143. Finder JD, Engman CL, Kagan VE, et al. Bronchoalveolar lavage fluid from patients with cystic fibrosis have diminished levels of reduced glutathione [abstract]. *Ped Pulm* 2003; 239 Suppl. 25: A158
144. Hull J, Vervaart P, Grimwood K, et al. Pulmonary oxidative stress response in young children with cystic fibrosis. *Thorax* 1997; 52: 557-60
145. Tirouvanziam R. CFTR, glutathione, and neutrophil function [online]. Available from URL: <http://63.193.197.190/Subsets/tirouv/index.htm> [Accessed 1999 Nov 13]
146. Neeffes VM, Evelo CT, Baars LG, et al. Erythrocyte glutathione S transferase as a marker of oxidative stress at birth. *Arch Dis Child Fetal Neonatal Ed* 1999 Sep; 81 (2): F130-3
147. Brown RK, Wyatt H, Price JF, et al. Pulmonary dysfunction in cystic fibrosis is associated with oxidative stress. *Eur Respir J* 1996; 9: 334-9
148. Roum JH, Buhl R, McElvaney NG, et al. Systemic deficiency of glutathione in cystic fibrosis. *J Appl Physiol* 1993 Dec; 75 (6): 2419-24
149. Tirouvanziam RM, Tjioe I, Moss RB, et al. Multiparameter study of CF blood and lung leukocytes using 3-laser, 11-color cytometry. *Pediatr Pulmonol* 2001 Oct; 22: 271-2
150. Abraham EH, Sterling KM, Kim RJH, et al. Erythrocyte membrane ATP binding cassette (ABC) proteins: MRP1 and CFTR as well as CD39 (ectoapyrase) involved in RBC ATP transport and elevated blood plasma ATP of cystic fibrosis. *Blood Cells Mol Dis* 2001 Jan-Feb; 27 (1): 165-80
151. Dominguez C, Gartner S, Linan S, et al. Enhanced oxidative damage in cystic fibrosis patients. *Biofactors* 1998; 8 (1-2): 149-53
152. Lloyd-Still JD, Ganther HE. Selenium and glutathione peroxidase levels in cystic fibrosis. *Pediatrics* 1980; 65: 1010-2
153. Carmagnol F, Sinet PM, Lenoir G, et al. Absence of modifications of the enzyme defense system against oxygen toxicity in cystic fibrosis. *Pediatr Res* 1983; 17: 181-2
154. Konstan MW, Berger M. Current understanding of the inflammatory process in cystic fibrosis: onset and etiology. *Pediatr Pulmonol* 1997 Aug; 24 (2): 137-42
155. Tirouvanziam R, Khazaal I, Peault B. Primary inflammation in human cystic fibrosis small airways. *Am J Physiol Lung Cell Mol Physiol* 2002 Aug; 283 (2): L445-51
156. Tirouvanziam R, de Bentzmann S, Hubeau C, et al. Inflammation and infection in naive human cystic fibrosis airway grafts. *Am J Respir Cell Mol Biol* 2000 Aug; 23 (2): 121-7
157. Escotte S, Laplace V, Benoit S, et al. Basal KC production in tracheal surface liquid of CF mice engrafted in nude mice [abstract]. *Pediatr Pulmonol* 2001 Oct; 22: 227
158. Lancellotti L, D'Orazio C, Mastella G, et al. Deficiency of vitamins E and A in cystic fibrosis is independent of pancreatic function and current enzyme and vitamin supplementation. *Eur J Pediatr* 1996 Apr; 155 (4): 281-5
159. Portal BC, Richard MJ, Faure HS, et al. Altered antioxidant status and increased lipid peroxidation in children with cystic fibrosis. *Am J Clin Nutr* 1995 Apr; 61 (4): 843-7
160. Winklhofer-Roob BM, Ellemunter H, Fruhwirth M, et al. Plasma vitamin C concentrations in patients with cystic fibrosis: evidence of associations with lung inflammation. *Am J Clin Nutr* 1997 Jun; 65 (6): 1858-66
161. Wood LG, Fitzgerald DA, Gibson PG, et al. Oxidative stress in cystic fibrosis: dietary and metabolic factors. *Am Coll Nutr* 2001 Apr; 20 (2 Suppl.): 157-65
162. Madarasi A, Lugassi A, Greiner E, et al. Antioxidant status in patients with cystic fibrosis. *Ann Nutr Metab* 2000; 44 (5-6): 207-11
163. Isabelle D, Puget M, Bellon G, et al. Antioxidant and peroxidative status in 312 patients with cystic fibrosis. *Pediatr Pulmonol* 2002 Oct; 24: 339
164. Birrer P, McElvaney NG, Rudeberg A, et al. Protease-antiprotease imbalance in the lungs of children with cystic fibrosis. *Am J Respir Crit Care Med* 1994 Jul; 150 (1): 207-13
165. Griese M, Birrer P, Demirsoy A. Pulmonary surfactant in cystic fibrosis. *Eur Respir J* 1997 Sep; 10 (9): 1983-8
166. DiMango E, Tabibi S, Ratner A, et al. Activation of NF-kappaB by adherent *Pseudomonas aeruginosa* in normal cystic fibrosis respiratory epithelial cells. *J Clin Invest* 1998; 101 (11): 2598-605
167. Weber A, Nguyen B, Bryan R, et al. Effects of CFTR mutations on epithelial cytokine expression [abstract]. *Pediatr Pulmonol* 1999 Sep; Suppl. 19: 309-10
168. Elborn JS, Cordon SM, Western PJ, et al. Tumor necrosis-factor-alpha, resting energy expenditure and cachexia in cystic fibrosis. *Clin Sci (Colch)* 1993 Nov; 85 (5): 563-8
169. Pfeffer KD, Huecksteadt TP, Hoidal JR. Expression and regulation of tumor necrosis factor in macrophages from cystic fibrosis patients. *Am J Respir Cell Mol Biol* 1993 Nov; 9 (5): 511-9
170. Greally P, Hussain MJ, Vergani D, et al. Serum interleukin-1 alpha and soluble interleukin-2 receptor concentrations in cystic fibrosis. *Arch Dis Child* 1993 Jun; 68 (6): 785-7
171. Wilmott RW, Frenzke M, Kociela V, et al. Plasma interleukin-1 alpha and beta, tumor necrosis factor-alpha, and lipopolysaccharide concentrations during pulmonary exacerbations of cystic fibrosis. *Pediatr Pulmonol* 1994 Jul; 18 (1): 21-7
172. Lundberg JO, Nordvall SL, Weitzberg E, et al. Exhaled nitric oxide in paediatric asthma and cystic fibrosis. *Arch Dis Child* 1996; 75: 323-6
173. Kroesbergen A, Jobsis Q, Bel EH, et al. Flow-dependency of exhaled nitric oxide in children with asthma and cystic fibrosis. *Eur Respir J* 1999 Oct; 14 (4): 871-5
174. Grasmann H, Michler E, Wallot M, et al. Decreased concentration of exhaled nitric oxide (NO) in patients with cystic fibrosis. *Pediatr Pulmonol* 1997; 24: 173-7
175. Balfour-Lynn IM, Laverty A, Dinwiddie R. Reduced upper airway nitric oxide in cystic fibrosis. *Arch Dis Child* 1996; 75: 319-22
176. Dotsch J, Demirakca S, Terbrack HG, et al. Airway nitric oxide in asthmatic children and patients with cystic fibrosis. *Eur Respir J* 1996; 9: 2537-40
177. Thomas SR, Kharitonov SA, Scott SF, et al. Nasal and exhaled nitric oxide is reduced in adult patients with cystic fibrosis and does not correlate with cystic fibrosis genotype. *Chest* 2000 Apr; 117 (4): 1085-9
178. Ho LP, Innes JA, Greening AP. Nitrite levels in breath condensate of patients with cystic fibrosis is elevated in contrast to exhaled nitric oxide. *Thorax* 1998 Aug; 53 (8): 680-4
179. Andersson C, Gaston B, Roomans G. S-Nitrosoglutathione induces functional deltaF508-CFTR in airway epithelial cells. *Biochem Biophys Res Commun* 2002 Sep 27; 297 (3): 552-7

180. Snyder AH, McPherson ME, Hunt JF, et al. Acute effects of aerosolized S-nitrosoglutathione in cystic fibrosis. *Am J Respir Crit Care Med* 2002 Apr 1; 165 (7): 922-6
181. Jungas T, Motta I, Duffieux F, et al. Glutathione levels and BAX activation during apoptosis due to oxidative stress in cells expressing wild-type and mutant cystic fibrosis transmembrane conductance regulator. *J Biol Chem* 2002 Aug 2; 277 (31): 27912-8
182. Hosseini Nia T, Kubow S, Grey V, et al. Inhibition of *Pseudomonas aeruginosa*-mediated mortality and weight loss via ingestion of whey treated by hyperbaric pressure [abstract]. *Pediatr Pulmonol* 2002 Oct; 24: 276
183. Buhl R, Vogelmeier C, Critenden M, et al. Augmentation of the glutathione in the fluid lining the epithelium of the lower respiratory tract by directly administering glutathione. *Proc Natl Acad Sci U S A* 1990 Jun; 87: 4063-7
184. Buhl R, Holroyd K, Borok Z, et al. Reversal of the glutathione deficiency in the lower respiratory tract of HIV seropositive individuals by glutathione aerosol therapy [abstract]. *Clin Res* 1990; 38 (2): 596A
185. Holroyd KJ, Buhl R, Borok Z, et al. Correction of glutathione deficiency in the lower respiratory tract of HIV seropositive individuals by glutathione aerosol treatment. *Thorax* 1993 Oct; 48 (10): 985-9
186. Borok Z, Buhl R, Grimes GJ, et al. Effect of glutathione aerosol on oxidant-antioxidant imbalance in idiopathic pulmonary fibrosis. *Lancet* 1991 Jul 17; 338: 215-6
187. Marrades RM, Roca J, Barbera JA, et al. Nebulized glutathione induces bronchoconstriction in patients with mild asthma. *Am J Respir Crit Care Med* 1997; 156: 425-30
188. Bernorio S, Pecis M, Zucchi A, et al. Glutathione in bronchial hyperresponsiveness. *J Aerosol Med* 1996; 9 (2): 207-13
189. Testa B, Mesolella M, Testa D, et al. Glutathione in the upper respiratory tract. *Ann Otol Rhinol Laryngol* 1995; 104: 117-9
190. Roum JH, Borok Z, McElvaney NG, et al. Glutathione aerosol suppresses lung epithelial surface inflammatory cell-derived oxidants in cystic fibrosis. *J Appl Physiol* 1999 Jul; 87 (1): 438-43
191. Bagnato GF, Gulli S, De Pasquale R, et al. Effect of inhaled glutathione on airway response to 'Fog' challenge in asthmatic patients. *Respiration* 1999 Nov-Dec; 66 (6): 518-21
192. Griese M, Ramakers J, Krasselt A, et al. Optimized aerosol delivery of glutathione (GSH) to patients with cystic fibrosis (CF) increases alveolar GSH concentration [abstract]. *Am J Respir Crit Care Med* 2003 Apr; 167 (7): A919
193. Griese M, Ramakers J, Krasselt A, et al. Improvement of alveolar glutathione and lung function but not oxidative state in cystic fibrosis. *Am J Respir Crit Care Med* 2004 Apr 1; 169 (7): 822-8
194. Bishop C, Hudson VM, Hilton SC, et al. Effect of inhaled buffered glutathione (GSH) on the clinical status of cystic fibrosis patients [abstract]. *Am J Respir Crit Care Med* 2003 Apr; 167 (7): A918
195. Bishop CT, Hudson VM, Hilton SC, Wilde C. A pilot study of the effect of inhaled buffered reduced glutathione on the clinical status of cystic fibrosis patients. *Chest*. In press
196. Aw TY, Wierzbicka G, Jones DP. Oral glutathione increases tissue glutathione in rats. *Chem Biol Interact* 1991; 80 (1): 89-97
197. Iantomasi T, Favilli F, Marraccini P, et al. Glutathione transport system in human small intestine epithelial cells. *Biochim Biophys Acta* 1997 Dec 4; 1330 (2): 274-83
198. Hunjan MK, Evered DF. Absorption of glutathione from the gastro-intestinal tract. *Biochim Biophys Acta* 1985 May 14; 815 (2): 184-8
199. Favilli F, Marraccini P, Iantomasi T, et al. Effect of orally administered glutathione on glutathione levels in some organs of rats: role of specific transporters. *Br J Nutr* 1997 Aug; 78 (2): 293-300
200. Hagen TM, Wierzbicka GT, Sillau AH, et al. Bioavailability of dietary glutathione: effect on plasma concentration. *Am J Physiol* 1990 Oct; 259 (4 Pt 1): G524-9
201. Gao L, Broughman JR, Iwamoto T, et al. Synthetic chloride channel restores glutathione secretion in cystic fibrosis airway epithelia. *Am J Physiol Lung Cell Mol Physiol* 2001 Jul; 281 (1): L24-30
202. Ciuchi E, Odetti P, Prando R. The effect of acute glutathione treatment on sorbitol level in erythrocytes from diabetic patients. *Diabetes Metab* 1997 Feb; 23 (1): 58-60
203. Sechi G, Deledda MG, Bua G, et al. Reduced intravenous glutathione in the treatment of early Parkinson's disease. *Prog Neuropsychopharmacol Biol Psychiatry* 1996 Oct; 20 (7): 1159-70
204. Visca AG. Adherence to treatment and self-prescription on new drugs: an emerging problem [abstract]. *Pediatr Pulmonol* 2002 Oct; 24: 347

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